

BIOSYNTHESIS AND OPTIMIZATION OF
METHYL 3-(3,5-DI-TERT-BUTYL-4-
HYDROXYPHENYL) PROPIONATE
PRODUCTION FROM OIL PALM FROND
JUICE BY *Ceratocystis fimbriata*

NANG NOR AZIMAH LONG NADZRI

MASTER OF SCIENCE

UNIVERSITI MALAYSIA PAHANG



SUPERVISOR'S DECLARATION

We hereby declare that We have checked this thesis and, in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science.

(Supervisor's Signature)

Full Name : DR MIOR AHMAD KHUSHAIRI MOHD ZAHARI

Position : SENIOR LECTURER

Date :

(Co-supervisor's Signature)

Full Name : PROF MADYA DR SAIFUL NIZAM TAJUDDIN

Position : ASSOCIATE PROFESSOR

Date :



STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

(Student's Signature)

Full Name : NANG NOR AZIMAH LONG NADZRI

ID Number : MKB15009

Date :

BIOSYNTHESIS AND OPTIMIZATION OF METHYL 3-(3,5-DI-TERT-BUTYL-
4-HYDROXYPHENYL) PROPIONATE PRODUCTION FROM OIL PALM
FROND JUICE BY *Ceratocystis fimbriata*

NANG NOR AZIMAH LONG NADZRI

Thesis submitted in fulfillment of the requirements
for the award of the degree of
Master of Science

Faculty of Chemical & Natural Resources Engineering
UNIVERSITI MALAYSIA PAHANG

AUGUST 2018

This page is entirely dedicated to.....

..... my lovely mother and father (Zakiah Bt Awang and Long Nadzri Bin Long Hussin), husband (Mohammad Firdaus Bin Abu Bakar), family and friends who have always been at my side and given me the encouragement and support that carries me through my study. Thanks for their never-ending love, support and care to me.....

(May ALLAH S.W.T. always be with all of you)

ACKNOWLEDGEMENTS

**In The Name of Allah, The Most Merciful, The Most Compassionate,
Peace and Blessings be Upon His Beloved Prophet.**

I am making uncounted praises to my Allah the Almighty who has guided me to remember Him at this time. I praised Him, for it is Him who has made this master study possible. Nothing is possible unless He made it possible.

Firstly, I would like to express my sincere gratitude to my advisor Dr Mior Ahmad Khushairi Mohd Zahari for the continuous support of my master study and related research, for his patience, motivation, doa' and immense knowledge. His guidance helped me in all the time of research and writing of this thesis. Besides my advisor, I would like to thank the rest of my thesis committee: Prof. Madya Dr Saiful Nizam Tajuddin for his insightful comments and encouragement, but also for the hard question which incited me to widen my research from various perspectives.

My sincere thanks also go to Bioaromatic's Lab, Universiti Malaysia Pahang and Chemical Engineering's Lab, Faculty of Chemical and Natural Resources Engineering, Universiti Malaysia Pahang because give me opportunity for access and using the laboratory and research facilities. Without the facilities support it would not be possible to conduct this research. I would also like to express my gratitude to Universiti Malaysia Pahang, which granted me the financial assistance under UMP Grant Scheme project no. RDU 150315 and PGRS160324.

I would like to thank the graduate students of Faculty of Chemical and Natural Resources Engineering, Universiti Malaysia Pahang for the stimulating discussions, for the sleepless nights we were working together before deadlines, and for all the fun we have had in the last two years (November 2015 until December 2017). Special thanks to all my friends for their friendship.

Last but not the least, to my husband and parents, I would like to say that without your sacrifice, encouragement and understandings I may not be able to complete this study. To my brothers and sisters, thank you for supporting me spiritually throughout the writing this thesis and my life in general especially for finance.

ABSTRAK

Di Malaysia, ladang kelapa sawit dan industri minyak sawit merupakan penyumbang utama kepada pembentukan sisa pertanian. Dari kajian terdahulu, penyelidikan telah dijalankan untuk mengkaji potensi penggunaan sisa pertanian dengan cekap. Dalam kajian ini, jus kelapa sawit (OPF) digunakan untuk menggantikan fungsi glukosa semasa proses penapaian. Jus OPF dilaporkan mengandungi gula boleh diperbaharui seperti glukosa, sukrosa dan fruktosa. Jus OPF dijangka boleh menangani isu-isu alam sekitar untuk menghasilkan sebatian organik mudah meruap (VOC) terutamanya metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat kerana OPF banyak terbuang sebagai biojisim dan mudah diperolehi di seluruh Malaysia. Kajian penggunaan dan pembangunan penghasilan metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat dari kulat semakin meningkat kerana ia boleh dihasilkan secara semula jadi tanpa sintesis kimia. *Ceratocystis fimbriata* adalah kulat yang mempunyai potensi untuk mensintesis ester, ia tumbuh dengan cepat dan menghasilkan pelbagai aroma (pic, nanas, pisang, sitrus dan ros) bergantung kepada keadaan persekitaran dan kultur yang digunakan. Tujuan utama kajian ini adalah untuk menyaring dan mengoptimumkan penghasilan metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat dengan menggunakan satu siri reka bentuk eksperimen dengan teknik pengekstrakan fasa pepejal ruang (HS-SPME) dengan menggunakan gas kromatografi-spektroskopi jisim (GC-MS) untuk memisahkan kawasan puncak relatif sebatian selepas proses penapaian. Pengoptimuman penghasilan metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat dipengaruhi oleh beberapa faktor semasa proses penapaian. Siri reka bentuk eksperimen digunakan untuk menyaring dan mengoptimumkan pengeluaran sebatian itu. Dalam kajian penyaringan, kaedah rekabentuk faktorial penuh 2^4 telah digunakan untuk mencari faktor-faktor penting yang mempengaruhi pengeluaran metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat, yang merupakan suhu inkubasi ($25\text{ }^{\circ}\text{C}$ - $35\text{ }^{\circ}\text{C}$), medium pH awal (pH4 - pH 8), kelajuan agitasi (100 rpm - 150 rpm) dan kepekatan glukosa (20 g/L - 30 g/L) dalam jus OPF. Respon dalam penapisan dipadankan dengan persamaan regresi linear berganda dan memperoleh korelasi ($R^2 = 0.8960$) antara data eksperimen dan data model. Reka bentuk komposit pusat (CCD) digunakan sebagai reka bentuk eksperimen dan model regresi polinomial dengan istilah kuadrat digunakan untuk menganalisis data eksperimen menggunakan analisis varians (ANOVA). Analisis ANOVA menunjukkan bahawa model sangat signifikan ($p < 0.0001$) untuk menghasilkan metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat. Respon tersebut dipadankan dengan persamaan polinomial urutan kedua dengan korelasi tinggi ($R^2 = 0.9598$) di antara nilai ujikaji dan nilai yang diramalkan. Keputusan proses pengoptimuman menunjukkan bahawa penghasilan propionat maksimum metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat diperolehi dalam keadaan medium pH awal (8), kelajuan agitasi (100 rpm) dan inkubasi suhu ($25\text{ }^{\circ}\text{C}$). Di bawah keadaan optimum ini, pengeluaran metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat tertinggi didapati apabila masa penahanan adalah pada 32.80 minit dan kawasan puncak relatif 0.29% kawasan kromatogram dengan menggunakan GC-SPME. Kesimpulan kajian ini telah memberikan garis panduan yang signifikan dan kefahaman awal bagi penghasilan sebatian metil 3- (3,5-di-tert-butil-4-hidroksifenil) propionat dengan menggunakan jus OPF sebagai substrat tunggal, boleh diperolehi dan mampan oleh *C. fimbriata* pada skala yang lebih besar pada masa hadapan.

ABSTRACT

In Malaysia, oil palm plantations and the palm oil industries were the main contributors to the generation of agricultural waste. From previous studies, researchers have identified the potential of utilizing the agricultural waste efficiently. In this research, oil palm frond (OPF) juice was used to replace the function of glucose during fermentation. OPF juice is reported to contain renewable sugars such as glucose, sucrose and fructose. OPF juice is expected to address the environmental issues to produce volatile organic compounds (VOCs) especially for production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate as OPF is abundantly available as a biomass and easily available throughout Malaysia. The utilization and development of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production fungus have been of increasing interest as they are naturally produced without chemical synthesis. *Ceratocystis fimbriata* is a fungus which has the potential for synthesizing esters, it grows quickly and produces a variety of aromas (peach, pineapple, banana, citrus and rose) depending on the strain and culture conditions. The aim of this study was to screen and optimize methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate using a series of experimental design by a head space-solid phase micro extraction (HS-SPME) technique combined with gas chromatography-mass spectroscopy (GC-MS) was used to separate the relative peak area of the compound during the fermentation. Optimization of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production are affected by several factors during the period of fermentation. Series of experimental designs were applied to screen and optimize the production of the compound. In the screening study, 2^4 full factorial design were used to find significant factors affecting production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate, which are incubation temperature (25 °C-35 °C), initial pH medium (pH4, pH8), agitation speed (100 rpm, 150 rpm) and concentration of glucose (20 g/L, 30 g/L) in OPF juice. The responses in screening were fitted with a multiple linear regression equation and obtained a correlation ($R^2 = 0.8960$) between the experimental data and model data. Then central composite design (CCD) was applied as the experimental design and a polynomial regression model with quadratic term was used to analyze the experimental data using analysis of variance (ANOVA). ANOVA analysis showed that the model was very significant ($p < 0.0001$) for the methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production. The responses were fitted with the second order polynomial equation with high correlation ($R^2 = 0.9598$) between the observed and predicted values. The results of optimization process showed that a maximum methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production was obtained under the condition of initial pH medium (8), agitation speed (100 rpm) and incubation temperature (25°C). Under these optimized conditions, the highest 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production was obtained when the column retention time was 32.80 minutes and the relative peak area was 0.29 % of chromatogram area by using GC-SPME. As a conclusion, this study provides a significant guideline and basic of understanding for the production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate compound at larger scale using OPF juice as sole, renewable and sustainable substrate by *C. fimbriata* in the near future.

TABLE OF CONTENT

DECLARATION

TITLE PAGE

ACKNOWLEDGEMENTS **ii**

ABSTRAK **iii**

ABSTRACT **iv**

TABLE OF CONTENT **v**

LIST OF TABLES **ix**

LIST OF FIGURES **xi**

LIST OF SYMBOLS **xiii**

LIST OF ABBREVIATIONS **xiv**

CHAPTER 1 INTRODUCTION **1**

1.1 Background 1

1.2 Problem statements 3

1.3 Objectives of study 5

1.4 Scope of the study 6

CHAPTER 2 LITERATURE REVIEW **7**

2.1 Introduction 7

2.2 Volatile organic compounds and microorganisms 7

2.3 *Ceratocystis fimbriata* 9

2.4 Volatile organic compounds by biosynthesis 10

2.4.1 Alcohol 10

2.4.2	Ester	11
2.4.3	Methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	11
2.4.4	Aldehyde	13
2.5	Analytical method for analysis volatile organic compounds	14
2.6	Production of VOCs from low cost (waste-based substrate)	16
2.6.1	Production of VOCs from cassava wastewater	17
2.6.2	Production of VOCs from apple pomace	17
2.6.3	Production of VOCs from soybean	17
2.6.4	Production of VOCs from amaranth grain	18
2.6.5	Production of VOCs from OPF juice	18
2.7	Renewable sugar from OPF	21
2.8	Characterization of OPF and OPF juice as fermentation substrate	22
2.9	Factors affecting production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	25
2.9.1	Incubation temperature	25
2.9.2	Initial pH medium	26
2.9.3	Agitation speed	26
2.9.4	Glucose concentration	26
2.10	Design of experiment	27
2.10.1	Factorial analysis	27
2.10.2	Optimization design	28
CHAPTER 3 METHODOLOGY		30
3.1	Overall research methodology	30
3.2	Preparation of OPF juice	32
3.3	Sugar content	33

3.4	Fungus strain	33
3.5	Growth and production medium for <i>C. fimbriata</i>	33
3.6	Fermentation procedure	35
3.7	Fermentation procedure for screening and optimization	36
3.8	Cell dry weight (CDW) measurement	36
3.9	Determination of residual sugar	37
3.10	Analytical procedure	37
3.11	Screening process using full factorial analysis	37
3.12	Optimization process using central composite design	39
3.13	Validation experiments	41
CHAPTER 4 RESULTS AND DISCUSSION		42
4.1	Preliminary study	42
4.1.1	Cell biomass and growth profile of <i>C. fimbriata</i>	43
4.1.2	VOCs production	47
4.1.3	Summary for preliminary study	52
4.2	Factorial analysis for production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	52
4.2.1	Model fitting	54
4.2.2	Analysis of variance (Anova)	57
4.2.3	Effects on main factors on methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	59
4.2.4	Validation of experiment	63
4.2.5	Summary for screening	64
4.3	Optimization for production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	64
4.3.1	Analysis of Variance (ANOVA)	66

4.3.2	Parametric interaction effect	69
4.3.3	Validation of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate optimization	74
4.3.4	Optimization of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	75
4.3.5	Summary for the optimization	77
CHAPTER 5 CONCLUSION AND RECOMMENDATION		78
5.1	Conclusion	78
5.2	Recommendation	79
REFERENCES		81
APPENDIX A HPLC STANDARD CURVE		100
APPENDIX B SCREENING AND OPTIMIZATION USING GC-MS SPME ANALYSIS		102
APPENDIX C FACTORIAL ANALYSIS		108
APPENDIX D CENTRAL COMPOSITE DESIGN		110
APPENDIX E LIST OF PUBLICATION		113

LIST OF TABLES

Table 2.1	Physical properties of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	13
Table 2.2	Sources and types of oil palm residues	20
Table 2.3	Amount of sugars contained in the OPF juice from different section of fresh oil palm frond	22
Table 2.4	Nutrient and metallic elements in OPF and OPF juice *	23
Table 2.5	Amino acids content in OPF juice	24
Table 3.1	Basal growth medium for <i>C. fimbriata</i>	34
Table 3.2	MSM for <i>C. fimbriata</i>	35
Table 3.3	Factors and levels used in the 2 ⁴ factorial design study	38
Table 3.4	Experimental design matrix for screening	39
Table 3.5	Experimental range and levels of the factors	40
Table 3.6	Experimental design matrix using RSM with CCD and response for methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	40
Table 4.1	Amount of sugar contain in OPF juice by using HPLC	43
Table 4.2	Volatile organic compounds produced by <i>C. fimbriata</i> using OPF juice as the sole carbon source	50
Table 4.3	Volatile organic compounds produced by <i>C. fimbriata</i> using OPF juice as the sole carbon source for 8 days of fermentation (summarized from Table 4.2)	52
Table 4.4	Experimental result for production of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	53
Table 4.5	The percentage contribution of each main factors and their interaction.	55
Table 4.6	Analysis of variance (ANOVA) analysis for 2 ⁴ full factorial design (FFD)	58
Table 4.7	Suggested best condition for factors in methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	63
Table 4.8	Comparison between predicted and experimental value for best condition	64
Table 4.9	Experimental design using RSM with CCD and response for methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	65
Table 4.10	ANOVA for the experimental results of CCD quadratic model	67
Table 4.11	Conditions for optimizing methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate factors	75

Table 4.12 Comparison between predicted and experimental value for optimum condition

75

LIST OF FIGURES

Figure 2.1	Methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate	13
Figure 2.2	The distribution of oil palm plantations in Malaysia	19
Figure 3.1	Research methodology for production methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate from OPF juice	31
Figure 3.2	(a) Freshly OPF without leaves, (b) Sugarcane pressing machine used in this study, (c) OPF after pressing, (d) Fresh OPF juice after pressing	32
Figure 4.1	(a) <i>C. fimbriata</i> after 7 days on PDA plate at incubation temperature of 30 °C, (b) <i>C. fimbriata</i> under microscope had cylindrical endoconidia (c) <i>C. fimbriata</i> had wide-mouth endoconidiophore with emerging doliiform endoconidium	44
Figure 4.2	(a) Seed culture of <i>C. fimbriata</i> in basal growth medium for the nine days on a rotary shaker at (150 rpm) in an aerobic condition, (b) The mycellium cell on the first day of fermentation, (c) The second day of fermentation, (d) third day of fermentation, (e) on the fourth day of fermentation	45
Figure 4.3	Glucose consumption profile for eight day of fermentation period by <i>C. fimbriata</i> in OPF juice supplemented with MSM.	46
Figure 4.4	Biomass production of <i>C. fimbriata</i> in OPF juices supplemented with MSM as a sole substrate for 8 days of fermentation	46
Figure 4.5	(GC-MS) with (SPME) analysis produced 9 peaks on the fourth day of fermentation. Peak (1) retention time at 8.661, peak (2) retention time at 10.421, peak (3) retention time at 12.732 were the alcohol group; peak (6) retention time at 22.643 and peak (9) retention time at 32.858 were the ester group; peak (4) retention time at 15.604 was the aldehyde group; peak (5) retention time at 18.107 was fatty acid; peak (7) retention time at 24.044 was the phenol group; and peak (8) retention time at 32.174 was the ketone group	49
Figure 4.6	Pareto chart of effects of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	57
Figure 4.7	The initial pH medium effect and agitation speed (rpm) on the methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	61
Figure 4.8	The glucose concentration in OPF juice effect and temperature (°C) on the methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	62
Figure 4.9	The glucose concentration in OPF juice effect and agitation speed (rpm) on the methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	63
Figure 4.10	Actual vs predicted response of methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate response	68

Figure 4.11	Residual plot for methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate response	69
Figure 4.12	Response surface and contour plot showing the effect of initial pH medium and agitation speed on methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	73
Figure 4.13	Response surface and contour plot showing the effect of incubation temperature and agitation speed on methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	73
Figure 4.14	Response surface and contour plot showing the effect of incubation temperature and initial pH medium on methyl 3-(3,5-di-tert-butyl-4-hydroxyphenyl) propionate production	74

LIST OF SYMBOLS

mL	milliliter
L	Liter
kg	kilogram
g	gram
°C	temperature
rpm	Rotation per minute
g/L	Gram per Liter
mL/min	Milliliter per minute
spores/mL	Spores per milliliter

LIST OF ABBREVIATIONS

GC-MS	Gas chromatography mass spectrometry
SPME	Solid phase micro extraction
HPLC	High performance liquid chromatography
ANOVA	Analysis of variance
<i>C. fimbriata</i>	<i>Ceratocystis fimbriata</i>
VOCs	Volatile organic compounds
CCD	Central composite design
DOE	Design of experiments
OPF	Oil palm frond
OPT	Oil palm trunk
EFB	Empty fruit bunches
PKC	Palm kernel cake
POME	Palm oil mill effluent
PPF	Palm press fibre
FFB	Fresh fruit bunches
MARDI	Malaysian Agricultural Research and Development Institute
CDW	Cell dry weight
FFD	Full factorial design
RSM	Research surface methodology
PDMS	Polydimethylsiloxane
CAR	Carboxen
DVB	Divinylbenzene
MPOB	Malaysian palm oil board
PKS	Palm kernel shells
MF	Mesocarp fibres
NaOH	Sodium hydroxide
HCL	Hydrochloric acid
MSM	Mineral salt medium

$(\text{NH}_4)_2\text{SO}_4$	Ammonium sulfate
KH_2PO_4	Monopotassium phosphate
$\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$	Calcium nitrate tetrahydrate
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	Magnesium sulfate heptahydrate
$\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$	Iron(III) nitrate nonahydrate
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	Zinc sulfate heptahydrate
$(\text{MnSO}_4 \cdot 4\text{H}_2\text{O})$	Manganese(II) sulfate tetrahydrate
MSM	Mineral salt medium
ATCC	American type of culture collection
PDA	Potato dextrose agar

REFERENCES

- Abdullah, R., & Wahid, M. B. (2010). World palm oil supply, demand, price and prospects: focus on Malaysian and Indonesian palm oil industry. *Malaysian Palm Oil Board Press, Malaysia*.
- Abnisa, F., Arami-Niya, A., Daud, W. W., Sahu, J., & Noor, I. (2013). Utilization of oil palm tree residues to produce bio-oil and bio-char via pyrolysis. *Energy conversion and management*, 76, 1073-1082.
- Agency, M. I. (2011). National Biomass Strategy 2020: New wealth creation for Malaysia's palm oil industry. Retrieved from <https://innovation.my/>
- Alam, M. Z., Mamun, A. A., Qudsieh, I. Y., Muyibi, S. A., Salleh, H. M., & Omar, N. M. (2009). Solid state bioconversion of oil palm empty fruit bunches for cellulase enzyme production using a rotary drum bioreactor. *Biochemical Engineering Journal*, 46(1), 61-64.
- Alam, M. Z., Muyibi, S. A., Mansor, M. F., & Wahid, R. (2007). Activated carbons derived from oil palm empty-fruit bunches: Application to environmental problems. *Journal of Environmental Sciences*, 19(1), 103-108.
- Albuquerque, P. M., Koch, F., Trossini, T. G., Esposito, E., & Ninow, J. L. (2006). Production of *Rhizopus oligosporus* Protein by Solid-State Fermentation of Apple Pomace. *Brazilian Archives of Biology and Technology*, 49(I), 91.
- Andersen, T. B., Cozzi, F., & Simonsen, H. T. (2015). Optimization of biochemical screening methods for volatile and unstable sesquiterpenoids using HS-SPME-GC-MS. *Chromatography*, 2(2), 277-292.
- Anderson, M., Whitcomb, P., Kraber, S., & Adams, W. (2009). Handbook for experimenters. *Stat-Ease, Inc*.
- Antony, J., & Roy, R. K. (1999). Improving the process quality using statistical design of experiments: a case study. *Quality Assurance: Good Practice, Regulation, and Law*, 6(2), 87-95.
- Ariffin, H., Abdullah, N., Umi Kalsom, M., Shirai, Y., & Hassan, M. (2006). Production and characterization of cellulase by *Bacillus pumilus* EB3. *Int. J. Eng. Technol*, 3(1), 47-53.
- Asma, Mahanim S, Zulkafli H, Othman S, & Mori. (2010). *Malaysia oil palm biomass. Forest Research Institute Malaysia*,. Paper presented at the Regional workshop on UNEP/DTIE/IETC in collaboration with GEC,, Osaka Japan.
- Bach, S. S., Bassard, J.-É., Andersen-Ranberg, J., Møldrup, M. E., Simonsen, H. T., & Hamberger, B. (2014). High-throughput testing of terpenoid biosynthesis candidate genes using transient expression in *Nicotiana benthamiana*. *Plant Isoprenoids: Methods and Protocols*, 245-255.

- Baharuddin, A. S., Hock, L. S., Yusof, M., Rahman, N. A. A., Shah, U., Hassan, M. A., Shirai, Y. (2010). Effects of palm oil mill effluent (POME) anaerobic sludge from 500 m³ of closed anaerobic methane digested tank on pressed-shredded empty fruit bunch (EFB) composting process. *African Journal of Biotechnology*, 9(16), 2427-2436.
- Bandaiphet, C., & Prasertsan, P. (2006). Effect of aeration and agitation rates and scale-up on oxygen transfer coefficient, k_{La} in exopolysaccharide production from *Enterobacter cloacae* WD7. *Carbohydrate polymers*, 66(2), 216-228.
- Baranski, J. R. (2008). Method for the preparation of a hydroxyalkyl hindered phenolic antioxidant: Google Patents.
- Baranyi, J., & Roberts, T. A. (1994). A dynamic approach to predicting bacterial growth in food. *International journal of food microbiology*, 23(3-4), 277-294.
- Barka, N., Abdennouri, M., Boussaoud, A., Galadi, A., Baâlala, M., Bensitel, M., Sadiq, M. (2014). Full factorial experimental design applied to oxalic acid photocatalytic degradation in TiO₂ aqueous suspension. *Arabian Journal of Chemistry*, 7(5), 752-757.
- Baş, D., & Boyacı, İ. H. (2007). Modeling and optimization I: Usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836-845.
- Bauer, K., Garbe, D., & Surburg, H. (2008). *Common fragrance and flavor materials: preparation, properties and uses*: John Wiley & Sons.
- Bengaly, K., Liang, J., Jelan, Z., Ho, Y., & Ong, H. (2010). Utilization of steam-processed oil palm (*Elaeis guineensis*) frond by ruminants in Malaysia: investigations for nitrogen supplementation. *African Journal of Agricultural Research*, 5(16), 2131-2136.
- Berger, R., Neuhäuser, K., & Drawert, F. (1987). Biotechnological production of flavor compounds: III. High productivity fermentation of volatile flavors using a strain of *Ischnoderma benzoinum*. *Biotechnology and bioengineering*, 30(8), 987-990.
- Berovič, M., & Ostroveršnik, H. (1997). Production of *Aspergillus niger* pectolytic enzymes by solid state bioprocessing of apple pomace. *Journal of Biotechnology*, 53(1), 47-53.
- Bertoldo, M., & Ciardelli, F. (2004). Water extraction and degradation of a sterically hindered phenolic antioxidant in polypropylene films. *Polymer*, 45(26), 8751-8759.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965-977.

- Bhalla, T., & Joshi, M. (1994). Protein enrichment of apple pomace by co-culture of cellulolytic moulds and yeasts. *World journal of microbiology and biotechnology*, 10(1), 116-117.
- Bigelis, R. (1992). Flavor metabolites and enzymes from filamentous fungi. *Food technology (USA)*.
- Bluemke, W., & Schrader, J. (2001). Integrated bioprocess for enhanced production of natural flavors and fragrances by *Ceratocystis moniliformis*. *Biomolecular Engineering*, 17(4), 137-142.
- Bramorski, A., Soccol, C. R., Christen, P., & Revah, S. (1998). Fruity aroma production by *Ceratocystis fimbriata* in solid cultures from agro-industrial wastes. *Revista de Microbiologia*, 29(3).
- Braunegg, G., Lefebvre, G., & Genser, K. F. (1998). Polyhydroxyalkanoates, biopolyesters from renewable resources: physiological and engineering aspects. *Journal of Biotechnology*, 65(2), 127-161.
- Breheret, S., Talou, T., Rapior, S., & Bessière, J.-M. (1997). Monoterpenes in the aromas of fresh wild mushrooms (Basidiomycetes). *Journal of agricultural and food chemistry*, 45(3), 831-836.
- Brigido, B. M. (2000). Produção de compostos voláteis de aroma por novas linhagens de *Neurospora* sp.
- Brown, A., & Hammond, J. (2003). Flavour control in small-scale beer fermentations. *Food and bioproducts processing*, 81(1), 40-49.
- Burdock, G. A. (2016). *Fenaroli's handbook of flavor ingredients*: CRC press.
- Bushman, Z. (2015). Ester Production in Fermentation. Retrieved from <https://www.gastrograph.com/blogs/gastronexus/ester-production-in-fermentation.html>
- Campillo, N., Vinas, P., López-García, I., Aguinaga, N., & Hernández-Córdoba, M. (2004). Purge-and-trap capillary gas chromatography with atomic emission detection for volatile halogenated organic compounds determination in waters and beverages. *Journal of Chromatography A*, 1035(1), 1-8.
- Carvalho, F., Garrote, G., Parajó, J. C., Pereira, H., & Gírio, F. M. (2005). Kinetic Modeling of Brewerywaste Spent Grain Autohydrolysis. *Biotechnology progress*, 21(1), 233-243.
- Che Maail, C. M. H., Ariffin, H., Hassan, M. A., Shah, U. K. M., & Shirai, Y. (2014). Oil palm frond juice as future fermentation substrate: a feasibility study. *BioMed Research International*, 2014.
- Cheetham, P. S. (1997). Combining the technical push and the business pull for natural flavours *Biotechnology of aroma compounds* (pp. 1-49): Springer.

- Chiron, N., & Michelot, D. (2005). Odeurs des champignons: chimie et rôle dans les interactions biotiques-une revue. *Cryptogamie. Mycologie*, 26(4), 299-364.
- Cho, I., Namgung, H.-J., Choi, H.-K., & Kim, Y.-S. (2008). Volatiles and key odorants in the pileus and stipe of pine-mushroom (*Tricholoma matsutake* Sing.). *Food Chemistry*, 106(1), 71-76.
- Christen, P., Meza, J., & Revah, S. (1997). Fruity aroma production in solid state fermentation by *Ceratocystis fimbriata*: influence of the substrate type and the presence of precursors. *Mycological Research*, 101(8), 911-919.
- Christen, P., & Raimbault, M. (1991). Optimization of culture medium for aroma production by *Ceratocystis fimbriata*. *Biotechnology letters*, 13(7), 521-526.
- Christen, P., Villegas, E., & Revah, S. (1994). Growth and aroma production by *Ceratocystis fimbriata* in various fermentation media. *Biotechnology letters*, 16(11), 1183-1188.
- Chung, H. Y. (1999). Volatile components in fermented soybean (*Glycine max*) curds. *Journal of agricultural and food chemistry*, 47(7), 2690-2696.
- Chung, H. Y., Fung, P. K., & Kim, J.-S. (2005). Aroma impact components in commercial plain sufu. *Journal of agricultural and food chemistry*, 53(5), 1684-1691.
- Ciganek, M., Pisarikova, B., & Zraly, Z. (2007). Determination of volatile organic compounds in the crude and heat treated amaranth samples. *Veterinarni medicina-praha-*, 52(3), 111.
- Claeson, A. S., & Sunesson, A. L. (2005). Identification using versatile sampling and analytical methods of volatile compounds from *Streptomyces albidoflavus* grown on four humid building materials and one synthetic medium. *Indoor Air*, 15(s9), 41-47.
- Clark, D. S., & Blanch, H. W. (1997). *Biochemical engineering*: CRC Press.
- Comi, G., Romano, P., Cocolin, L., & Fiore, C. (2001). Characterization of *Kloeckera apiculata* strains from the Friuli region in Northern Italy. *World journal of microbiology and biotechnology*, 17(4), 391-394.
- Correa, A. D., Jokl, L., & Carlsson, R. (1986). Chemical constituents, in vitro protein digestibility, and presence of antinutritional substances in amaranth grains. *Archivos latinoamericanos de nutricion*, 36(2), 319-326.
- Dahlan, I. (2000). Oil palm frond, a feed for herbivores. *Asian Australasian Journal of Animal Sciences*, 13, 300-303.
- Dalsenter, F. D. H., Viccini, G., Barga, M. C., Mitchell, D. A., & Krieger, N. (2005). A mathematical model describing the effect of temperature variations on the kinetics of microbial growth in solid-state culture. *Process Biochemistry*, 40(2), 801-807.

- Damasceno, S., Cereda, M., Pastore, G., & Oliveira, J. (2003). Production of volatile compounds by *Geotrichum fragrans* using cassava wastewater as substrate. *Process Biochemistry*, 39(4), 411-414.
- Darah, I., & Ibrahim, C. (1996). Effect of agitation on production of lignin-degrading enzymes by *Phanerochaete chrysosporium* grown in shake-flask cultures. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 4(3), 174-182.
- Darah, I., Sumathi, G., Jain, K., & Lim, S. (2011). Influence of agitation speed on tannase production and morphology of *Aspergillus niger* FETL FT3 in submerged fermentation. *Applied biochemistry and biotechnology*, 165(7-8), 1682-1690.
- Devrajan, A., Joshi, V. K., Gupta, K., Sheikher, C., & Lal, B. B. (2004). Evaluation of apple pomace based reconstituted feed in rats after solid state fermentation and ethanol recovery. *Brazilian Archives of Biology and Technology*, 47(1), 93-106.
- Dewulf, J., Van Langenhove, H., & Wittmann, G. (2002). Analysis of volatile organic compounds using gas chromatography. *TrAC Trends in Analytical Chemistry*, 21(9), 637-646.
- Diaconescu, R. M., Grigoriu, A.-M., Luca, C., & Georgescu, P. (2011). Study on the response surface modelling by central composite design and optimization of paper nanocoating. *Revista de Chimie, București*, 62(5), 522.
- Dragone, G., Silva, D. P., & e Silva, J. B. d. A. (2004). Factors influencing ethanol production rates at high-gravity brewing. *LWT-Food Science and Technology*, 37(7), 797-802.
- Dufour, J., Malcorps, P., & Silcock, P. (2008). 21 Control of ester synthesis during brewery fermentation. *Brewing yeast fermentation performance*.
- Eiceman, G. A., Gardea-Torresdey, J., Overton, E., Carney, K., & Dorman, F. (2004). Gas chromatography. *Analytical chemistry*, 76(12), 3387-3394.
- Engelbrecht, C. J. B., & Harrington, T. C. (2005). Intersterility, morphology and taxonomy of *Ceratocystis fimbriata* on sweet potato, cacao and sycamore. *Mycologia*, 97(1), 57-69.
- Evans, A. Mohagheghi, J. Hamilton, & Zhang, M. (2002). *Effect of corn stover hydrolysate and temperature on fermentation performance of selected yeast strains*. Paper presented at the Proceedings of the 24th Biotechnology for Fuels and Chemicals Symposium,, Gatlinburg, TN, USA.
- Fang, H., Zhao, C., & Song, X.-Y. (2010). Optimization of enzymatic hydrolysis of steam-exploded corn stover by two approaches: Response surface methodology or using cellulase from mixed cultures of *Trichoderma reesei* RUT-C30 and *Aspergillus niger* NL02. *Bioresource technology*, 101(11), 4111-4119.
- Farbood, M. I. (1991). Micro-organisms as a novel source of flavour compounds: Portland Press Limited.

- Favela-Torres, E., Volke-Sepúlveda, T., & Viniegra-González, G. (2006). Production of Hydrolytic Depolymerising Pectinases. *Food Technology & Biotechnology*, 44(2).
- Fernández, M., Ubeda, J., & Briones, A. (2000). Typing of non-Saccharomyces yeasts with enzymatic activities of interest in wine-making. *International journal of food microbiology*, 59(1), 29-36.
- Foo, L. Y., & Lu, Y. (1999). Isolation and identification of procyanidins in apple pomace. *Food Chemistry*, 64(4), 511-518.
- Fraatz, M. A., & Zorn, H. (2011). Fungal flavours *Industrial Applications* (pp. 249-268): Springer.
- Fujita, E. M., Harshfield, G., & Sheetz, L. (2003). Performance audits and laboratory comparisons for SCOS97-NARSTO measurements of speciated volatile organic compounds. *Atmospheric environment*, 37, 135-147.
- Fung, D.-R., Chuang, J.-J., Huang, Z.-J., & Chen, C.-Y. (2014). Method for making hindered phenolic antioxidant: Google Patents.
- Gatto, V. J., Elnagar, H. Y., Cheng, C. H., & Adams, J. R. (2011). Preparation of sterically hindered hydroxyphenylcarboxylic acid esters: Google Patents.
- Giavasis, I., Harvey, L. M., & McNeil, B. (2006). The effect of agitation and aeration on the synthesis and molecular weight of gellan in batch cultures of *Sphingomonas paucimobilis*. *Enzyme and Microbial Technology*, 38(1-2), 101-108.
- Górecki, T., Yu, X., & Pawliszyn, J. (1999). Theory of analyte extraction by selected porous polymer SPME fibres. *Analyst*, 124(5), 643-649.
- Grigelmo-Miguel, N., & Martín-Belloso, O. (1999). Comparison of dietary fibre from by-products of processing fruits and greens and from cereals. *LWT-Food Science and Technology*, 32(8), 503-508.
- Hamad, H. O., Alma, M. H., Ismael, H. M., & Gocer, A. (2014). The effect of some sugars on the growth of *Aspergillus niger*. *Kahramanmaraş Sütçü İmam Üniversitesi Doğa Bilimleri Dergisi*, 17(4), 7-11.
- Hassan, Ishida, M., Shukri, I. M., & Tajuddin, Z. A. (1996). Oil-palm fronds as a roughage feed source for ruminants in Malaysia. *Extension bulletin. Food and Fertilizer Technology Center for the Asian and Pacific Region*, 1-8.
- Hassan, Shirai, Y., Umeki, H., Yamazumi, H., Jin, S., Yamamoto, S., Hashimoto, K. (1997). Acetic Acid Separation from Anaerobically Treated Palm Oil Mill Effluent by Ion Exchange Resins for the Production of Polyhydroxyalkanoate by *Alcaligenes eutrophus*. *Bioscience, biotechnology, and biochemistry*, 61(9), 1465-1468.

- Hellen, H., Hakola, H., Pirjola, L., Laurila, T., & Pystynen, K.-H. (2006). Ambient air concentrations, source profiles, and source apportionment of 71 different C2–C10 volatile organic compounds in urban and residential areas of Finland. *Environmental science & technology*, 40(1), 103-108.
- Hernández-Carbajal, G., Rutiaga-Quñones, O. M., Pérez-Silva, A., Saucedo-Castañeda, G., Medeiros, A., Soccol, C. R., & Soto-Cruz, N. Ó. (2013). Screening of native yeast from *Agave duranguensis* fermentation for isoamyl acetate production. *Brazilian Archives of Biology and Technology*, 56(3), 357-363.
- Hettenhaus, J. R. (1998). Ethanol fermentation strains: present and future requirements for biomass to ethanol commercialization
- Hunt, J. (1956). Taxonomy of the genus *Ceratocystis*. *Lloydia*, 19, 1-59.
- Hussin, M. H., Rahim, A. A., Ibrahim, M. N. M., & Brosse, N. (2013). Physicochemical characterization of alkaline and ethanol organosolv lignins from oil palm (*Elaeis guineensis*) fronds as phenol substitutes for green material applications. *Industrial crops and products*, 49, 23-32.
- Imeri, A., Flores, R., Elias, L., & Bressani, R. (1987). Effect of processing and amino acids supplementation on the protein quality of amaranth (*Amaranthus caudatus*). *Archivos latinoamericanos de nutricion*, 37(1), 161-173.
- Jafari, A., Sarrafzadeh, M., Alemzadeh, I., & Vosoughi, M. (2007). Effect of stirrer speed and aeration rate on the production of glucose oxidase by *Aspergillus niger*. *Journal of Biological Sciences*, 7(2), 270-275.
- Johnson, J. A., Harrington, T. C., & Engelbrecht, C. (2005). Phylogeny and taxonomy of the North American clade of the *Ceratocystis fimbriata* complex. *Mycologia*, 97(5), 1067-1092.
- Kalaichelvan, P. (2012). Production and optimization of Pectinase from *Bacillus* sp. MFW7 using cassava waste. *Asian Journal of Plant Science and Research*, 2(3), 369-375.
- Kataoka, H., & Saito, K. (2011). Recent advances in SPME techniques in biomedical analysis. *Journal of pharmaceutical and biomedical analysis*, 54(5), 926-950.
- Kawamoto, H., Mohamed, W. Z., Shukur, N. I. M., Ali, M. S. M., Ismail, Y., & Oshio, S. (2001). Palatability, digestibility and voluntary intake of processed oil palm fronds in cattle. *Japan Agricultural Research Quarterly: JARQ*, 35(3), 195-200.
- Khanna, S., & Srivastava, A. K. (2005). Recent advances in microbial polyhydroxyalkanoates. *Process Biochemistry*, 40(2), 607-619.
- Kim, I., & Han, J.-I. (2012). Optimization of alkaline pretreatment conditions for enhancing glucose yield of rice straw by response surface methodology. *biomass and bioenergy*, 46, 210-217.

- Kłosowski, G., & Czupryński, B. (2006). Kinetics of acetals and esters formation during alcoholic fermentation of various starchy raw materials with application of yeasts *Saccharomyces cerevisiae*. *Journal of Food Engineering*, 72(3), 242-246.
- Kobayashi, M., Shimizu, H., & Shioya, S. (2008). Beer volatile compounds and their application to low-malt beer fermentation. *Journal of bioscience and bioengineering*, 106(4), 317-323.
- Koller, M., Atlić, A., Dias, M., Reiterer, A., & Brauneegg, G. (2010). Microbial PHA production from waste raw materials *Plastics from bacteria* (pp. 85-119): Springer.
- Korpi, A., Järnberg, J., & Pasanen, A.-L. (2009). Microbial volatile organic compounds. *Critical reviews in toxicology*, 39(2), 139-193.
- Kosugi, A., Tanaka, R., Magara, K., Murata, Y., Arai, T., Sulaiman, O., Yusof, M. N. M. (2010). Ethanol and lactic acid production using sap squeezed from old oil palm trunks felled for replanting. *Journal of bioscience and bioengineering*, 110(3), 322-325.
- Kot-Wasik, A., Dębska, J., & Namieśnik, J. (2004). Monitoring of organic pollutants in coastal waters of the Gulf of Gdańsk, Southern Baltic. *Marine pollution bulletin*, 49(3), 264-276.
- Krishna, & Karanth, N. (2002). Response Surface Modeling of Lipase-Catalyzed Isoamyl Propionate Synthesis. *Journal of food science*, 67(1), 32-36.
- Ku, K.-L., Chen, T.-P., & Chiou, R.-Y. (2000). Apparatus used for small-scale volatile extraction from ethanol-supplemented low-salt miso and GC– MS characterization of the extracted flavors. *Journal of agricultural and food chemistry*, 48(8), 3507-3511.
- Kunkee, R., & Amerine, M. (1970). Yeasts in wine-making. *The yeasts*, 3, 5-72.
- Lanza, E., Ko, K. H., & Palmer, J. K. (1976). Aroma production by cultures of *Ceratocystis moniliformis*. *Journal of agricultural and food chemistry*, 24(6), 1247-1250.
- Latrasse, A., Dameron, P., Hassani, H., & Staron, T. (1987). Formation of a fruity aroma by *Geotrichum candidum*. *Sciences des Aliments*, 7, 637-645.
- Lazic, Z. R. (2006). *Design of experiments in chemical engineering: a practical guide*: John Wiley & Sons.
- Lee, Azizan, M. N., & Sudesh, K. (2004). Effects of culture conditions on the composition of poly (3-hydroxybutyrate-co-4-hydroxybutyrate) synthesized by *Comamonas acidovorans*. *Polymer Degradation and Stability*, 84(1), 129-134.

- Lee, Teong, K., & Bhatia, S. (2010). Hot compressed water pretreatment of oil palm fronds to enhance glucose recovery for production of second generation bioethanol. *Bioresource technology*, 101(19), 7362-7367.
- Lee, C., & Abdul Halim, F. (2014). Oil Palm Fronds Juice: A potential Feedstock for Bioethanol Production. *International Journal of Scientific and Research Publications*, 4(12), 520-526.
- Leejeerajumnean, A., Duckham, S. C., Owens, J. D., & Ames, J. M. (2001). Volatile compounds in bacillus-fermented soybeans. *Journal of the Science of Food and Agriculture*, 81(5), 525-529.
- Li, X., Wang, Z.-G., Chen, H.-H., & Liu, S.-G. (2014). The antioxidant methyl 3-(3, 5-di-tert-butyl-4-hydroxyphenyl) propionate. *Acta Crystallographica Section C: Structural Chemistry*, 70(11), 1050-1053.
- Lin, Q., Li, H., Ren, J., Deng, A., Li, W., Liu, C., & Sun, R. (2017). Production of xylooligosaccharides by microwave-induced, organic acid-catalyzed hydrolysis of different xylan-type hemicelluloses: Optimization by response surface methodology. *Carbohydrate polymers*, 157, 214-225.
- Lomascolo, A., Stentelaire, C., Asther, M., & Lesage-Meessen, L. (1999). Basidiomycetes as new biotechnological tools to generate natural aromatic flavours for the food industry. *Trends in Biotechnology*, 17(7), 282-289.
- Loo, C.-Y., & Sudesh, K. (2007). Biosynthesis and native granule characteristics of poly (3-hydroxybutyrate-co-3-hydroxyvalerate) in *Delftia acidovorans*. *International journal of biological macromolecules*, 40(5), 466-471.
- Lu, Y., & Foo, L. Y. (2000). Antioxidant and radical scavenging activities of polyphenols from apple pomace. *Food Chemistry*, 68(1), 81-85.
- Mahlia, T., Abdulmuin, M., Alamsyah, T., & Mukhlisien, D. (2001). An alternative energy source from palm wastes industry for Malaysia and Indonesia. *Energy conversion and management*, 42(18), 2109-2118.
- Mangani, F., Maione, M., & Palma, P. (2003). 4 GC-MS Analysis of Halocarbons in the Environment. *Advances in chromatography*, 42, 139-240.
- Mantzouridou, F., Roukas, T., & Kotzekidou, P. (2002). Effect of the aeration rate and agitation speed on β -carotene production and morphology of *Blakeslea trispora* in a stirred tank reactor: mathematical modeling. *Biochemical Engineering Journal*, 10(2), 123-135.
- Martendal, E., Budziak, D., & Carasek, E. (2007). Application of fractional factorial experimental and Box-Behnken designs for optimization of single-drop microextraction of 2, 4, 6-trichloroanisole and 2, 4, 6-tribromoanisole from wine samples. *Journal of Chromatography A*, 1148(2), 131-136.

- Martin, A. M., & Bailey, V. I. (1985). Growth of *Agaricus campestris* NRRL 2334 in the form of pellets. *Applied and Environmental Microbiology*, 49(6), 1502-1506.
- Mason, R. L., Gunst, R. F., & Hess, J. L. (2003). *Statistical design and analysis of experiments: with applications to engineering and science* (Vol. 474): John Wiley & Sons.
- Masoodi, F., Sharma, B., & Chauhan, G. (2002). Use of apple pomace as a source of dietary fiber in cakes. *Plant Foods for Human Nutrition (Formerly Qualitas Plantarum)*, 57(2), 121-128.
- Matich, A. J., Rowan, D. D., & Guenther, C. (2008). Deceit and deception in volatile analysis. *Chemistry in New Zealand*, 73, 88-91.
- McMeekin, T., Olley, J., Ratkowsky, D., & Ross, T. (2002). Predictive microbiology: towards the interface and beyond. *International journal of food microbiology*, 73(2), 395-407.
- Medeiros, Christen, P., Roussos, S., Gern, J. C., & Soccol, C. R. (2003). Coffee residues as substrates for aroma production by *Ceratocystis fimbriata* in solid state fermentation. *Brazilian Journal of Microbiology*, 34(3), 245-248.
- Medeiros, Pandey, A., Christen, P., Fontoura, P. S., de Freitas, R. J., & Soccol, C. R. (2001). Aroma compounds produced by *Kluyveromyces marxianus* in solid state fermentation on a packed bed column bioreactor. *World journal of microbiology and biotechnology*, 17(8), 767-771.
- Medeiros, Pandey, A., Freitas, R. J., Christen, P., & Soccol, C. R. (2000). Optimization of the production of aroma compounds by *Kluyveromyces marxianus* in solid-state fermentation using factorial design and response surface methodology. *Biochemical Engineering Journal*, 6(1), 33-39.
- Medeiros, Pandey, A., Vandenberghe, L. P., Pastore, G. M., & Soccol, C. R. (2006). Production and recovery of aroma compounds produced by solid-state fermentation using different adsorbents. *Food Technology and Biotechnology*, 44(1), 47-51.
- Mee, R. (2009). *A comprehensive guide to factorial two-level experimentation*: Springer Science & Business Media.
- Merfort, I. (2002). Review of the analytical techniques for sesquiterpenes and sesquiterpene lactones. *Journal of Chromatography A*, 967(1), 115-130.
- Messens, W., Verluyten, J., Leroy, F., & De Vuyst, L. (2003). Modelling growth and bacteriocin production by *Lactobacillus curvatus* LTH 1174 in response to temperature and pH values used for European sausage fermentation processes. *International journal of food microbiology*, 81(1), 41-52.
- Montgomery, D. C. (2017). *Design and analysis of experiments*: John Wiley & Sons.

- Mori, Y., Kiuchi, K., & Tabei, H. (1983). Flavor components of miso: basic fraction. *Agricultural and biological chemistry*, 47(7), 1487-1492.
- Morimoto, M., Atsuko, M., Atif, A., Ngan, M., Fakhru'l-Razi, A., Iyuke, S., & Bakir, A. (2004). Biological production of hydrogen from glucose by natural anaerobic microflora. *International Journal of Hydrogen Energy*, 29(7), 709-713.
- MPOB. (2010). Foreword from the Chairman of the Malaysian Palm Oil Board. Retrieved 15 May 2017 from www.mpob.gov.my
- MPOB. (2011a). Annual and Forecast of Crude Palm Oil Production (Tonnes) 2010 & 2011. Retrieved 3 May 2017 from econ.mpob.gov.my/economy/EID_web.htm
- MPOB. (2011b). Oil Palm Planted Areas as at September 2011. Retrieved 18 May 2017 from econ.mpob.gov.my/economy/area/Area_category.pdf
- Mumtaz, T., Abd-Aziz, S., Rahman, N., Yee, P. L., Shirai, Y., & Hassan, M. (2008). Pilot-scale recovery of low molecular weight organic acids from anaerobically treated palm oil mill effluent (POME) with energy integrated system. *African Journal of Biotechnology*, 7(21).
- Mumtaz, T., Yahaya, N. A., Abd-Aziz, S., Yee, P. L., Shirai, Y., & Hassan, M. A. (2010). Turning waste to wealth-biodegradable plastics polyhydroxyalkanoates from palm oil mill effluent—a Malaysian perspective. *Journal of Cleaner Production*, 18(14), 1393-1402.
- Nasrah, N. S. M., Zahari, M. A. K. M., Masngut, N., & Ariffin, H. (2017). *Statistical Optimization for Biobutanol Production by Clostridium acetobutylicum ATCC 824 from Oil Palm Frond (OPF) Juice Using Response Surface Methodology*. Paper presented at the MATEC Web of Conferences.
- Nath, A., & Chattopadhyay, P. (2007). Optimization of oven toasting for improving crispness and other quality attributes of ready to eat potato-soy snack using response surface methodology. *Journal of Food Engineering*, 80(4), 1282-1292.
- Ngadi, M., & Correia, L. (1992a). Kinetics of solid-state ethanol fermentation from apple pomace. *Journal of Food Engineering*, 17(2), 97-116.
- Ngadi, M., & Correia, L. (1992b). Solid state ethanol fermentation of apple pomace as affected by moisture and bioreactor mixing speed. *Journal of food science*, 57(3), 667-670.
- Niinemets, Ü., Loreto, F., & Reichstein, M. (2004). Physiological and physicochemical controls on foliar volatile organic compound emissions. *Trends in plant science*, 9(4), 180-186.
- Oliveira, L., Guimarães, L., Ferreira, M., Nunes, A., Pimenta, L., & Alfenas, A. (2015). Aggressiveness, cultural characteristics and genetic variation of *Ceratocystis fimbriata* on *Eucalyptus* spp. *Forest Pathology*, 45(6), 505-514.

- Omar, R., Idris, A., Yunus, R., Khalid, K., & Isma, M. A. (2011). Characterization of empty fruit bunch for microwave-assisted pyrolysis. *Fuel*, 90(4), 1536-1544.
- Ooi, Z. X., Ismail, H., Bakar, A. A., & Teoh, Y. P. (2014). A review on recycling ash derived from *Elaeis Guineensis* by-product. *BioResources*, 9(4), 7926-7940.
- Ortíz-Castro, R., Contreras-Cornejo, H. A., Macías-Rodríguez, L., & López-Bucio, J. (2009). The role of microbial signals in plant growth and development. *Plant signaling & behavior*, 4(8), 701-712.
- Paganini, C., Nogueira, A., Silva, N. C., & Wosiacki, G. (2005). Utilization of apple pomace for ethanol production and food fiber obtainment. *Ciência e Agrotecnologia*, 29(6), 1231-1238.
- Pagans, E., Font, X., & Sánchez, A. (2006). Emission of volatile organic compounds from composting of different solid wastes: abatement by biofiltration. *Journal of hazardous materials*, 131(1), 179-186.
- Palmqvist, E., & Hågerdal, B. (2000). Fermentation of lignocellulosic hydrolysates. II: inhibitors and mechanisms of inhibition. *Bioresource technology*, 74(1), 25-33.
- Pandey, A., Soccol, C. R., Nigam, P., Soccol, V. T., Vandenberghe, L. P., & Mohan, R. (2000). Biotechnological potential of agro-industrial residues. II: cassava bagasse. *Bioresource technology*, 74(1), 81-87.
- Pastore, Park, Y., & Min, D. (1994). Production of fruity aroma by *Neurospora* from beiju. *Mycological Research*, 98(11), 1300-1302.
- Pastore, Sato, H. H., Yang, T.-S., Park, Y. K., & Min, D. B. (1994). Production of fruity aroma by newly isolated yeast. *Biotechnology letters*, 16(4), 389-392.
- Paul, J. S., Tiwari, K., & Jadhav, S. (2015). Long term preservation of commercial important fungi in glycerol at 4 C. *International Journal of Biological Chemistry*, 9(2), 79-85.
- Paulin, Harrington, T. C., & McNew, D. (2002). Phylogenetic and taxonomic evaluation of *Chalara*, *Chalaropsis*, and *Thielaviopsis* anamorphs associated with *Ceratocystis*. *Mycologia*, 94(1), 62-72.
- Pawliszyn, J. (2000). Theory of solid-phase microextraction. *Journal of chromatographic science*, 38(7), 270-278.
- Peddie, H. A. (1990). Ester formation in brewery fermentations. *Journal of the Institute of Brewing*, 96(5), 327-331.
- Pellati, F., Prencipe, F. P., & Benvenuti, S. (2013). Headspace solid-phase microextraction-gas chromatography-mass spectrometry characterization of propolis volatile compounds. *Journal of pharmaceutical and biomedical analysis*, 84, 103-111.

- Petronilho, S., Coimbra, M. A., & Rocha, S. M. (2014). A critical review on extraction techniques and gas chromatography based determination of grapevine derived sesquiterpenes. *Analytica chimica acta*, 846, 8-35.
- Popiel, S., & Sankowska, M. (2011). Determination of chemical warfare agents and related compounds in environmental samples by solid-phase microextraction with gas chromatography. *Journal of Chromatography A*, 1218(47), 8457-8479.
- Porcel, E. R., López, J. C., Pérez, J. S., Sevilla, J. F., & Chisti, Y. (2005). Effects of pellet morphology on broth rheology in fermentations of *Aspergillus terreus*. *Biochemical Engineering Journal*, 26(2), 139-144.
- Purwanto, L., Ibrahim, D., & Sudrajat, H. (2009). Effect of agitation speed on morphological changes in *Aspergillus niger* hyphae during production of tannase. *World J. Chem*, 4(1), 34-38.
- Qiao, D., Hu, B., Gan, D., Sun, Y., Ye, H., & Zeng, X. (2009). Extraction optimized by using response surface methodology, purification and preliminary characterization of polysaccharides from *Hyriopsis cumingii*. *Carbohydrate polymers*, 76(3), 422-429.
- Rahman, S., Choudhury, J., & Ahmad, A. (2006). Production of xylose from oil palm empty fruit bunch fiber using sulfuric acid. *Biochemical Engineering Journal*, 30(1), 97-103.
- Ramos-Jeunehomme, C. (1991). *Why is ester formation in brewery fermentations yeast strain dependent?* Paper presented at the Proceedings of the 23rd European Brewery Convention Congress.
- Rasat, M. S. M., Wahab, R., Sulaiman, O., Moktar, J., Mohamed, A., Tabet, T. A., & Khalid, I. (2011). Properties of composite boards from oil palm frond agricultural waste. *BioResources*, 6(4), 4389-4403.
- Rizk, M., Abdel-Rahman, T., & Metwally, H. (2007). Factors affecting growth and antifungal activity of some *Streptomyces* species against *Candida albicans*. *International Journal of food, agriculture and environment*, 5(3-4), 446-449.
- Rojas, V., Gil, J. V., Piñaga, F., & Manzanares, P. (2001). Studies on acetate ester production by non-*Saccharomyces* wine yeasts. *International journal of food microbiology*, 70(3), 283-289.
- Romano, P., & Marchese, R. (1998). Metabolic characterization of *Kloeckera apiculata* strains from star fruit fermentation. *Antonie van Leeuwenhoek*, 73(4), 321-325.
- Romano, P., Suzzi, G., Comi, G., Zironi, R., & Maifreni, M. (1997). Glycerol and other fermentation products of apiculate wine yeasts. *Journal of Applied Microbiology*, 82(5), 615-618.

- Romano, P., Suzzi, G., Domizio, P., & Fatichenti, F. (1997). Secondary products formation as a tool for discriminating non-Saccharomyces wine strains. *Antonie van Leeuwenhoek*, 71(3), 239-242.
- Rose, A. H., & Harrison, J. S. (2012). *The yeasts: Yeast technology* (Vol. 5): Elsevier.
- Roslan, A., Yee, P., Shah, U., Aziz, S., & Hassan, M. (2011). Production of bioethanol from rice straw using cellulase by local *Aspergillus* sp. *Int. J. Agric. Res*, 6(2), 188-193.
- Rosli, W. W., Law, K., Zainuddin, Z., & Asro, R. (2004). Effect of pulping variables on the characteristics of oil-palm frond-fiber. *Bioresource technology*, 93(3), 233-240.
- Rossi, S., Vandenberghe, L., Pereira, B., Gago, F., Rizzolo, J., Pandey, A., . . . Medeiros, A. (2009). Improving fruity aroma production by fungi in SSF using citric pulp. *Food research international*, 42(4), 484-486.
- Rozman, H., Kumar, R., Khalil, H. A., Abusamah, A., Lim, P., & Ismail, H. (1997). Preparation and properties of oil palm frond composite based on methacrylic silane and glycidyl methacrylate. *European polymer journal*, 33(3), 225-230.
- Rubiolo, P., Cagliero, C., Cordero, C., Liberto, E., Sgorbini, B., & Bicchi, C. (2014). Gas chromatography in the analysis of flavours and fragrances *Practical Gas Chromatography* (pp. 717-743): Springer.
- Rumbold, K., van Buijsen, H. J., Gray, V. M., van Groenestijn, J. W., Overkamp, K. M., Slomp, R. S., Punt, P. J. (2010). Microbial renewable feedstock utilization: a substrate-oriented approach. *Bioengineered bugs*, 1(5), 359-366.
- Sabiha, Noor, M. A. M., & Rosma, A. (2011). Effect of autohydrolysis and enzymatic treatment on oil palm (*Elaeis guineensis* Jacq.) frond fibres for xylose and xylooligosaccharides production. *Bioresource technology*, 102(2), 1234-1239.
- Salamatinia, B., Kamaruddin, A. H., & Abdullah, A. Z. (2010). Regeneration and reuse of spent NaOH-treated oil palm frond for copper and zinc removal from wastewater. *Chemical engineering journal*, 156(1), 141-145.
- Salmon, J., & Mauricio, J. (1994). Relationship between sugar uptake kinetics and total sugar consumption in different industrial *Saccharomyces cerevisiae* strains during alcoholic fermentation. *Biotechnology letters*, 16(1), 89-94.
- Sanchez, S., Bravo, V., Moya, A., Castro, E., & Camacho, F. (2004). Influence of temperature on the fermentation of D-xylose by *Pachysolen tannophilus* to produce ethanol and xylitol. *Process Biochemistry*, 39(6), 673-679.
- Santos, F., & Galceran, M. (2002). The application of gas chromatography to environmental analysis. *TrAC Trends in Analytical Chemistry*, 21(9), 672-685.

- Santosa, S. J. (2008). Palm oil boom in Indonesia: from plantation to downstream products and biodiesel. *CLEAN–Soil, Air, Water*, 36(5-6), 453-465.
- Saraji, M., & Ghani, M. (2015). Hollow fiber liquid–liquid–liquid microextraction followed by solid-phase microextraction and in situ derivatization for the determination of chlorophenols by gas chromatography-electron capture detection. *Journal of Chromatography A*, 1418, 45-53.
- Schindler, J. (1982). Terpenoids by microbial fermentation. *Industrial & Engineering Chemistry Product Research and Development*, 21(4), 537-539.
- Schrader, J. (2007). Microbial flavour production *Flavours and fragrances* (pp. 507-574): Springer.
- Seguchi, T., Tamura, K., Shimada, A., Sugimoto, M., & Kudoh, H. (2012). Mechanism of antioxidant interaction on polymer oxidation by thermal and radiation ageing. *Radiation Physics and Chemistry*, 81(11), 1747-1751.
- Senemaud, C. (1988). *Les champignons filamenteux producteurs d'aromes fruités: études de faisabilité sur substrats agro-industriels*. Dijon.
- Seth, M., & Chand, S. (2000). Biosynthesis of tannase and hydrolysis of tannins to gallic acid by *Aspergillus awamori*—optimisation of process parameters. *Process Biochemistry*, 36(1), 39-44.
- Shendell, D. G., Winer, A. M., Stock, T. H., Zhang, L., Zhang, J. J., Maberti, S., & Colome, S. D. (2004). Air concentrations of VOCs in portable and traditional classrooms: results of a pilot study in Los Angeles County. *Journal of Exposure Science and Environmental Epidemiology*, 14(1), 44.
- Shimizu, H., Tamura, S., Ishihara, Y., Shioya, S., & Suga, K. (1994). Control of molecular weight distribution and mole fraction in poly (-D (-)-3-hydroxyalkanoate)(PHA) production by *Mcaligenes eutrophus*. *Y. Doi*, 6, 389-394.
- Shirey, R. E., & Mindrup, R. F. (1999). SPME-adsorption versus absorption: which fiber is best for your application. *Supelco: Bellefonte, PA, USA*.
- Shojaosadati, S., & Babaeipour, V. (2002). Citric acid production from apple pomace in multi-layer packed bed solid-state bioreactor. *Process Biochemistry*, 37(8), 909-914.
- Shrikot, C., Sharma, N., & Sharma, S. (2004). Apple pomace: An alternative substrate for xylanase production by an alkalophilic *Bacillus macerans* by using solid-state fermentation. *Journal of Microbial World*, 6, 20-26.
- Shuit, S. H., Tan, K. T., Lee, K. T., & Kamaruddin, A. (2009). Oil palm biomass as a sustainable energy source: A Malaysian case study. *Energy*, 34(9), 1225-1235.

- Simon, M. J., Lagergren, E. S., Snyder, K. A., & Kenneth, A. (1997, October). Concrete mixture optimization using statistical mixture design methods. In *Proceedings of the PCI/FHWA international symposium on high performance concrete* (pp. 230-244).
- Soares, M., Christen, P., Pandey, A., & Soccol, C. R. (2000). Fruity flavour production by *Ceratocystis fimbriata* grown on coffee husk in solid-state fermentation. *Process Biochemistry*, 35(8), 857-861.
- Sobczuk, T. M., Camacho, F. G., Grima, E. M., & Chisti, Y. (2006). Effects of agitation on the microalgae *Phaeodactylum tricornutum* and *Porphyridium cruentum*. *Bioprocess and Biosystems Engineering*, 28(4), 243.
- Soccol, C. R., Medeiros, A. B., Vandenberghe, L. P., & Woiciechowski, A. L. (2007). Flavor Production by Solid and Liquid Fermentation. *Handbook of Food Products Manufacturing, 2 Volume Set*, 193.
- Sonyal, S. (2010). *Studies on pomegranate wilt complex*. University of Agricultural Sciences, Dharwad.
- Souza, Reyes-Garcés, N., Gómez-Ríos, G. A., Boyacı, E., Bojko, B., & Pawliszyn, J. (2015). A critical review of the state of the art of solid-phase microextraction of complex matrices III. Bioanalytical and clinical applications. *TrAC Trends in Analytical Chemistry*, 71, 249-264.
- Splivallo, R., Novero, M., Berteà, C. M., Bossi, S., & Bonfante, P. (2007). Truffle volatiles inhibit growth and induce an oxidative burst in *Arabidopsis thaliana*. *New Phytologist*, 175(3), 417-424.
- Steele, D. B., & Stowers, M. D. (1991). Techniques for selection of industrially important microorganisms. *Annual Reviews in Microbiology*, 45(1), 89-106.
- Strlič, M., Cigić, I. K., Rabin, I., Kolar, J., Pihlar, B., & Cassar, M. (2009). Autoxidation of lipids in parchment. *Polymer Degradation and Stability*, 94(6), 886-890.
- Sugawara, E. (1991). Change in aroma components of miso with aging. *Nippon Shokuhin Kogyo Gakkaishi*, 38(12), 1093-1097.
- Surburg, H., & Panten, J. (2016). *Common fragrance and flavor materials: preparation, properties and uses*: John Wiley & Sons.
- Talon, R., Chastagnac, C., Vergnais, L., Montel, M., & Berdagué, J. (1998). Production of esters by *Staphylococci*. *International journal of food microbiology*, 45(2), 143-150.
- Tan, H. T., Lee, K. T., & Mohamed, A. R. (2010). Optimizing ethanolic hot compressed water (EHCW) cooking as a pretreatment to glucose recovery for the production of fuel ethanol from oil palm frond (OPF). *Fuel Processing Technology*, 91(9), 1146-1151.

- Tan, H. T., Lee, K. T., & Mohamed, A. R. (2011). Pretreatment of lignocellulosic palm biomass using a solvent-ionic liquid [BMIM] Cl for glucose recovery: An optimisation study using response surface methodology. *Carbohydrate polymers*, 83(4), 1862-1868.
- Tan, J. P., Jahim, J. M., Harun, S., Wu, T. Y., & Mumtaz, T. (2016). Utilization of oil palm fronds as a sustainable carbon source in biorefineries. *International Journal of Hydrogen Energy*, 41(8), 4896-4906.
- Telford, J. K. (2007). A brief introduction to design of experiments. *Johns Hopkins apl technical digest*, 27(3), 224-232.
- Tirillini, B., Verdelli, G., Paolocci, F., Ciccio, P., & Frattoni, M. (2000). The volatile organic compounds from the mycelium of *Tuber borchii* Vitt. *Phytochemistry*, 55(8), 983-985.
- Tsurumi, R., Shiraishi, S., Ando, Y., Yanagida, M., & Takeda, K. (2001). Production of flavor compounds from apple pomace. *Journal-japanese society of food science and technology*, 48(8), 564-569.
- Uchiyama, S., Matsushima, E., Tokunaga, H., Otsubo, Y., & Ando, M. (2006). Determination of orthophthalaldehyde in air using 2, 4-dinitrophenylhydrazine-impregnated silica cartridge and high-performance liquid chromatography. *Journal of Chromatography A*, 1116(1), 165-171.
- Ujang, Z., Salmiati, S., & Salim, M. R. (2010). Microbial Biopolymerization Production from Palm Oil Mill Effluent (POME) *Biopolymers: InTech*.
- van Stee, L. L., Udo, A. T., & Bagheri, H. (2002). Gas chromatography with atomic emission detection: a powerful technique. *TrAC Trends in Analytical Chemistry*, 21(9), 618-626.
- Vandamme. (1996). Bacteria in front of the mirror-Biocosmetics via microbial synthesis. *Agro FOOD Industry Hi-Tech*, 7(4) 3-8).
- Vandamme. (2003). Bioflavours and fragrances via fungi and their enzymes. *Fungal Diversity*, 13, 153-166.
- Vendruscolo, F., Koch, F., de Oliveira Pitol, L., & Ninow, J. L. (2007). Produção de proteína unicelular a partir do bagaço de maçã utilizando fermentação em estado sólido. *Revista Brasileira de Tecnologia Agroindustrial*, 1(1).
- Venugopal, T., Jayachandra, K., & Appaiah, A. (2007). Effect of aeration on the production of endopectinase from coffee pulp by a novel thermophilic fungi, *Mycotypha* sp. strain no. AKM1801. *Biotechnology*, 6(2), 245-250.
- Verstrepen, K. J., Derdelinckx, G., Dufour, J.-P., Winderickx, J., Thevelein, J. M., Pretorius, I. S., & Delvaux, F. R. (2003). Flavor-active esters: adding fruitiness to beer. *Journal of bioscience and bioengineering*, 96(2), 110-118.

- Villas Bôas, S., & Esposito, E. (2000). Bioconversão do bagaço de maçã: enriquecimento nutricional utilizando fungos para produção de um alimento alternativo de alto valor agregado. *Revista de Biotecnologia, Brasília*, 1(14), 38-42.
- Wanrosli, W., Zainuddin, Z., Law, K., & Asro, R. (2007). Pulp from oil palm fronds by chemical processes. *Industrial crops and products*, 25(1), 89-94.
- Welsh. (1994). Overview of bioprocess flavor and fragrance production. *Bioprocess production of flavor, fragrance, and color ingredients*. Wiley, New York, 1-17.
- Welsh, Murray, W. D., Williams, R. E., & Katz, I. (1989). Microbiological and enzymatic production of flavor and fragrance chemicals. *Critical Reviews in Biotechnology*, 9(2), 105-169.
- Worrall, J., & Yang, C. S. (1992). Shiitake and oyster mushroom production on apple pomace and sawdust. *HortScience*, 27(10), 1131-1133.
- Wu, T., Mohammad, A. W., Jahim, J. M., & Anuar, N. (2006). Investigations on protease production by a wild-type *Aspergillus terreus* strain using diluted retentate of pre-filtered palm oil mill effluent (POME) as substrate. *Enzyme and Microbial Technology*, 39(6), 1223-1229.
- Yacob, S., Shirai, Y., Hassan, M. A., Wakisaka, M., & Subash, S. (2006). Start-up operation of semi-commercial closed anaerobic digester for palm oil mill effluent treatment. *Process Biochemistry*, 41(4), 962-964.
- Yamauchi, H., Obata, T., Amachi, T., & Hara, S. (1991). Production of characteristic odors by *Neurospora*. *Agricultural and biological chemistry*, 55(12), 3115-3116.
- Yee, P. L., Hassan, M. A., Shirai, Y., Wakisaka, M., & Karim, M. I. A. (2003). Continuous production of organic acids from palm oil mill effluent with sludge recycle by the freezing-thawing method. *Journal of chemical engineering of Japan*, 36(6), 707-710.
- Yegemova, S., Bakaikina, N. V., Kenessov, B., Koziel, J. A., & Nauryzbayev, M. (2015). Determination of 1-methyl-1H-1, 2, 4-triazole in soils contaminated by rocket fuel using solid-phase microextraction, isotope dilution and gas chromatography–mass spectrometry. *Talanta*, 143, 226-233.
- Yemiş, O., & Mazza, G. (2011). Acid-catalyzed conversion of xylose, xylan and straw into furfural by microwave-assisted reaction. *Bioresource technology*, 102(15), 7371-7378.
- Yusoff, S. (2006). Renewable energy from palm oil–innovation on effective utilization of waste. *Journal of Cleaner Production*, 14(1), 87-93.
- Zahari. (2013). *Oil Palm Frond Juice as a Novel and Renewable Substrate for the Production of Poly (3-hydroxybutyrate) Bioplastic*. Universiti Putra Malaysia.

- Zahari, Hassan, O. A., Wong, H., & Liang, J. (2003). Utilization of oil palm frond-based diets for beef and dairy production in Malaysia.
- Zahari, Zakaria, M. R., Ariffin, H., Mokhtar, M. N., Salihon, J., Shirai, Y., & Hassan, M. A. (2012). Renewable sugars from oil palm frond juice as an alternative novel fermentation feedstock for value-added products. *Bioresource technology*, 110, 566-571.
- Zain, S. M. S. M., Shaharudin, R., Kamaluddin, M. A., & Daud, S. F. (2017). Determination of hydrogen cyanide in residential ambient air using SPME coupled with GC-MS. *Atmospheric Pollution Research*.
- Zakaria, M. R., Tabatabaei, M., Ghazali, F. M., Abd-Aziz, S., Shirai, Y., & Hassan, M. A. (2010). Polyhydroxyalkanoate production from anaerobically treated palm oil mill effluent by new bacterial strain *Comamonas* sp. EB172. *World journal of microbiology and biotechnology*, 26(5), 767-774.
- Zawirska, W., Renata, Siwulski, M., & Mildner-Szkudlarz, S. (2009). Studies on the aroma of different species and strains of *Pleurotus* measured by GC/MS, sensory analysis and electronic nose. *Acta Scientiarum Polonorum Technologia Alimentaria*, 8(1), 47-61.
- Zhan, X., Zhang, Y.-H., Chen, D.-F., & Simonsen, H. T. (2014). Metabolic engineering of the moss *Physcomitrella patens* to produce the sesquiterpenoids patchoulol and α/β -santalene. *Frontiers in plant science*, 5.
- Zhang, Z., & Pawliszyn, J. (1993). Headspace solid-phase microextraction. *Analytical chemistry*, 65(14), 1843-1852.
- Zhang, Z., Yang, M. J., & Pawliszyn, J. (1994). Solid-phase microextraction. A solvent-free alternative for sample preparation. *Analytical chemistry*, 66(17), 844A-853A.
- Zheng, Z., & Shetty, K. (2000a). Enhancement of pea (*Pisum sativum*) seedling vigour and associated phenolic content by extracts of apple pomace fermented with *Trichoderma* spp. *Process Biochemistry*, 36(1), 79-84.
- Zheng, Z., & Shetty, K. (2000b). Solid state production of polygalacturonase by *Lentinus edodes* using fruit processing wastes. *Process Biochemistry*, 35(8), 825-830.
- Zygmunt, B., Zaborowska, A., & Namiesnik, J. (2007). Solid phase microextraction combined with gas chromatography-A powerful tool for the determination of chemical warfare agents and related compounds. *Current Organic Chemistry*, 11(3), 241-253.